Scheme III



the methyl group (Scheme I). F values⁹ of 37.2 and 37.5 (duplicate analyses) for the acetate from the first sample and 59.8 and 61.4 from the second indicated predominant S and R configuration, respectively, of the two acetate specimens. Since the degradation sequence involves one inversion of configuration in the cyanide displacement step to give acetonitrile, it follows that the first and the second 5-CH₃-H₄folate sample contained 44 \pm 1% ee R and $37 \pm 3\%$ ee S isomer, respectively (Scheme III). Hence the reduction of 5,10-CH2-H4folate had occurred stereoselectively, exhibiting a 2-3-fold preference for attack syn to C₆-H of the 5,10-CH₂-H₄folate molecule.

Samples of the chiral methyl-R and methyl-S 5-CH₃-H₄folate were then incubated with purified cobalamin-dependent methio-nine synthase from $E. \ coli^{10}$ in 0.1 M potassium phosphate buffer, pH 7.2, in the presence of 25 mM dithiothreitol, 50 µM aquocobalamin, 0.5 mM homocysteine, and 19 µM S-adenosylmethionine to give methionine in about 80% yield. Methionine was separated from residual 5-CH₃-H₄folate by passage over a column of AG1X8 (Bio-Rad) and then purified by HPLC on an ODS column equilibrated with 0.1 M ammonium acetate, pH 3.55. Methionine eluted at 5-7 min,¹¹ and residual ammonium acetate was removed by lyophilization. The purified methionine samples were degraded by the procedure of Arigoni and coworkers^{12,13} (Scheme II) to recover the methyl group as acetate for chirality analysis. This degradation sequence involves two inversions of the methyl group configuration, one in the displacement of methyl from S-methylmethionine by p-nitrothiobenzoate anion and another in the reaction of methyl p-nitrothiobenzoate with cyanide to give acetonitrile. Thus, the configuration of the acetate from the degradation directly reflects that of the methionine methyl group. F values of 42.5 and 44.2 (duplicate experiments) for the material derived from methyl-S-5-CH₃-H₄folate and 56.3 and 55.7 for that from methyl-R-5-CH₃-H₄folate indicated that the product of the enzyme reaction contained methyl groups of predominantly the same configuration as the substrate (Scheme III).

The results demonstrate that the cobalamin-dependent methionine synthase from E. coli transfers the methyl group of 5-CH₃-H₄folate stereoselectively to the sulfur of homocysteine to generate methionine with net retention of configuration. The observed steric course is consistent with the postulated mechanism of the reaction, which invokes two sequential transfers of the methyl group, one from 5-CH₃-H₄folate to cobalt to generate enzyme-bound methylcobalamin and a second from cobalt to sulfur to produce methionine. There is a substantial decrease in the chiral purity of the methyl group during the overall process. Since the degradation procedures for both CH₃-H₄folate and methionine are highly stereospecific,^{4,5,12,13} it follows that the enzymatic methyl transfer is accompanied by nearly 50% racemization. This observation may provide a clue for the further mechanistic analysis

37±3%eeS

F: 42.5, 44.2

23± 3% eeS

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of the reaction catalyzed by cobalamin-dependent methionine

B-Eliminations in Isonitriles

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The physical organic chemistry of the nitrile group has been the subject of intensive investigations¹ which have shown that is

synthase.

⁽¹⁰⁾ Frasca, V.; Matthews, R. G., unpublished method. The specific activity of the purified enzyme preparations used for these experiments was 0.86-3.72 IU (µmol min⁻¹ mg⁻¹), and the enzyme was about 25-80% pure as assessed by polyacrylamide gel electrophoresis in the presence of sodium dodecyl sulfate.

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Table I. Elimination in Isonitriles, G-CH₂CH₂-Z^a

substrate	<i>k</i> (⁻ CN) ^{<i>b</i>}	$k(Z^{-})^{b}$	activation ratio ^c -CN/-NC	products (%) from isonitriles			
				-CN	Z	$CH_2 = CH - Z$	CH ₂ =CH-NC
4a. CN-CH ₂ CH ₂ -OTs		1.3×10^{-1r}	3.8×10^4		82 ^d		64 ^e
4b. CN-CH ₂ CH ₂ -SO ₂ Ph	4.1×10^{-4}	$1.1 \times 10^{-4 \text{ h}}$	1.6×10^{4}	79⁄	658	15	7e
4c. CN-CH ₂ CH ₂ -SPh	$3.1 \times 10^{-5 k}$	$3.2 \times 10^{-6 h,k}$	3.1×10^{3}	13⁄	10	82 ^h	52'
4d. CN-CH ₂ CH ₂ -OPh	6.9×10^{-51}	$2.2 \times 10^{-6 h,l}$	4.3×10^{3}	27 ^f	25	74 ^h	7 2 ^j
4e, CN-CH ₂ CH ₂ -CN	3.0×10^{-2} s		р	m	88 ⁿ		
4g, CN-CH ₂ CH ₂ -Ph				q			

^aReactions in EtONa-EtOH at 25 °C unless otherwise stated. ^bdm³ mol⁻¹ s⁻¹. ^cData for nitriles from refs. 3 and 7. ^dAs NaOTs. ^eTrapped as 4c with PhS⁻-EtOH. ^fBy cyanide-sensitive electrode. ^gAs PhSO₂CH₂CH₂OEt. ^hBy UV spectroscopy. ^fBy trapping as 4e with KCN under phase-transfer conditions. ^fBy HPLC using product from 4a to calibrate and assuming 100% conversion. ^kExtrapolated from data from reactions >40 °C. 'Extrapolated from data from reactions >45 °C. "Detected but not accurately determined. "As NC-CH2CH2-OEt. "Substantial for KCN-Me₂SO reactions, see text. 9No CN after 18 h at 100 °C in M-EtONa-EtOH. 'Reactions followed by pH stat. 'Followed by appearance of NC-CH₂CH₂-OEt on GLC.

Scheme I

is a moderate carbanion stabilizer ($pK_a^{Me_2SO}$ CH₃CN 31.3, MeSO₂Me 31.1, MeCOMe 26.5)² and hence activates elimination reactions which involve expulsion of β -substituents (Scheme I).

Quantitative data³ for a large number of cyano-activated substrates (1a), with leaving groups Z = halogen, 'onium, SO₂Ph, SPh, OPh, etc.) have been obtained, and concerted or stepwise mechanisms were assigned on the basis of detritiation rates, calculated deprotonation rates, or exchange experiments.⁴ It was notable, that the nitrile group was itself never expelled in such eliminations in protic solvents.⁵ Because of the extraordinarily low nucleofugality⁶ of the cyano group, we have been prompted to examine the *isonitrile* group as both activator and nucleofuge in 1,2-eliminations.

Activation of elimination by the isonitrile group is compared (Table I) with that for the nitrile group and it can be seen that for all leaving groups, the nitrile group is a substantially superior activator of elimination. The mechanism for elimination of the nucleofuge, Z, is the same in both series; exchange of protium for deuterium adjacent to both functions in EtOD solutions is faster for substrates 4b-d than elimination, and we assign the $(E_1 cB)_R$ mechanism for these eliminations. For tosylate 4a, no such exchange can be detected, and for this substrate either the E_2 or the $(E_1 cB)_1$ mechanism operates. For the isomeric nitrile the $(E_1 cB)_1$ mechanism has been postulated.⁷ These results are all in accord with qualitative observations on isonitriles which suggest rather lower acidities than for nitriles.8

There are considerable differences in the nitrile/isonitrile activation ratios; as the leaving group becomes less inductive in the progression $SO_2Ph > OPh > SPh^9$ the ratio decreases. We attribute this trend to a lower sensitivity to induction of the preequilibrium formation of the carbanion in the isonitriles than in the nitriles. We are currently engaged in detritiation experiments to test this possibility.

The other striking feature of Table I is the elimination of the isonitrile group under activation by the group Z. Elimination of

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 CN^{-} from isonitriles, e.g., from m-NO₂C₆H₄CH₂C(NC)Ph₂, by the action of 50% NaOH/C₆H₆/n-Bu₄N⁺Br⁻ has been disclosed previously¹⁰ but no rate data were reported.

For sulfone 4b exchange experiments (EtOD) confirm that the $(E_1 cB)_R$ mechanism operates for the expulsion of both cyanide and sulfinate from this substrate. The calculated deprotonation rate constant together with the rate constant for the elimination of cyanide ion allows calculation of the rank (=nucleofugality = $\log k_{obsd} - \log k_{deprotonation} + 11)^6$ of the isonitrile group. Its rank of 3.3 places it well below leaving groups where C-halogen, C-S, C-O, or other C-N bonds are broken (NMeAc⁻ 6.6, NMe₂Ph 10.7).³ This value should obviously be compared with that of cyanide. Unfortunately, a direct comparison is impossible with the nitrile isomer of 4b; the sole of elimination product is benzenesulfinate ion.³ The rank (5.8) for isonitrile obtained from the nitrile-isonitrile 4e is not less than 10 units greater than for cyano in the same system.³

The failure to observe loss of cyanide ion from tosylate 4a is not surprising; the rapid elimination of tosylate ion must preclude the alternative pathway. Much more surprising is the loss of cyanide from the sulfide 4c and even more so from the phenyl ether 4d. We have established that for both substrates hydrogen-deuterium exchange with perdeuterioethanol as solvent is faster than elimination of either leaving group in both substrates. The isonitrile group is powerfully inductive ($\sigma_1 = 0.63$),¹¹ and this may account for deprotonation adjacent to the phenoxy group. Another remarkable example of a phenoxy-activated elimination has recently been reported¹² to occur in a platinum complex.

Isonitrile 4g is stable to ethoxide ethanol, and exchange occurs slowly adjacent to, and only adjacent to, the isonitrile group.

Behavior of the three possible nitrile-isonitrile isomers 4e, 4f, and 5 toward $K^{13}CN$ in Me_2SO-d_6 shown by ¹H and ¹³C NMR

$$\begin{array}{c} \text{CN-CH}_2\text{CH}_2\text{-NC} & \text{NC-CH}_2\text{CH}_2\text{-CN} \\ \textbf{4f} & \textbf{5} \end{array}$$

nicely illustrates the different behavior of the two functions. Isomer 4e shows conversion to 5, whereas 5 and 4f are inert under the conditions and so is isonitrile 4g. The lack of reactivity in 4g suggests that reaction of 4e proceeds not by substitution but by elimination-addition under activation by the cyano group. The nitrile group in 4e and 5 is too low ranked to be expelled and 4f is inert because the isonitrile group is neither a particularly good activating group nor a particularly good nucleofuge. Formation of an alkene from 4e under basic conditions has been referred to previously but no details are available.8

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